

**TRITERPENE GLYCOSIDES FROM *Astragalus* AND THEIR GENINS.
LXXXIX. ASKENDOSIDE H FROM *Astragalus taschkenticus***

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A new triterpene glycoside of the cycloartane series that was called askendoside H was isolated from roots of Astragalus taschkenticus Bunge (Leguminosae). Its structure was elucidated based on chemical transformations and spectral data. Askendoside H was a bisdesmoside of cycloorbigenin C, 23R,24R-cycloartan-3 β ,6 α ,16 β ,23,24,25-hexaol 3-O-[(α -L-arabinopyranosyl)(1 \rightarrow 2)- β -D-xylopyranoside] 23-O- β -D-glucopyranoside.

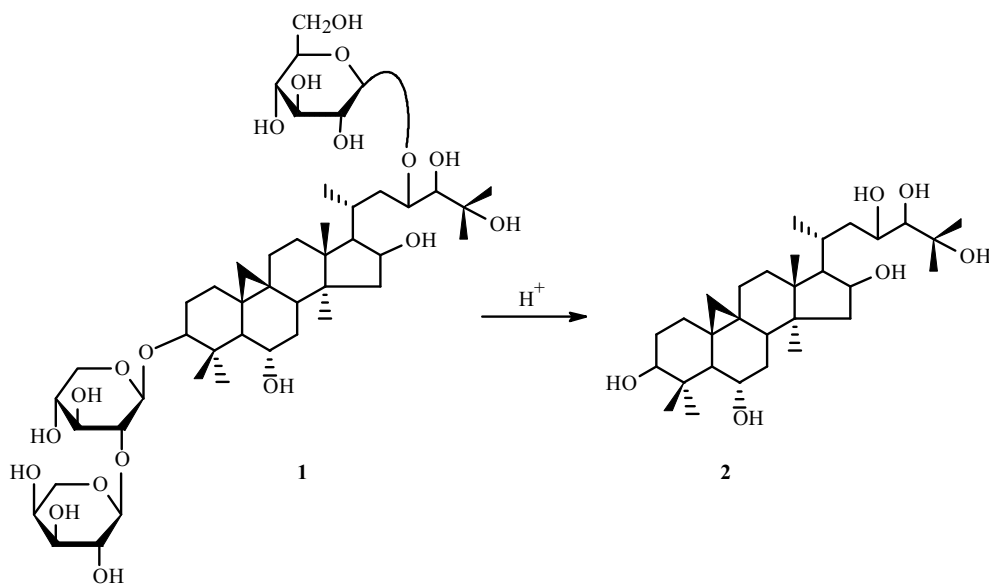
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In continuation of our chemical research on isoprenoids from plants of the genus *Astragalus* (Leguminosae), we isolated a new triterpene glycoside from roots of *A. taschkenticus* Bunge that we called askendoside H (**1**). Herein we provide proof of the structure of this glycoside.

The PMR spectrum of **1** exhibited at strong field δ 0.12 and 0.45 two ^1H resonances for an AX system that belonged to methylene of tetrasubstituted cyclopropane and resonances for seven methyls. This enabled us to classify **1** as a cycloartane series triterpenoid [1–4].

Acid hydrolysis of **1** and subsequent analysis of the carbohydrate part of the hydrolysate by paper chromatography (PC) identified taking into account biogenetic considerations D-xylose, D-glucose, and L-arabinose.

The ^{13}C NMR spectrum of **1** showed resonances for three anomeric C atoms at δ 105.61, 106.67, and 106.67 (Table 1), thereby showing that **1** contained D-xylose, D-glucose, and L-arabinose in a 1:1:1 ratio, i.e., the new glycoside was a trioside. The chemical shifts of the monosaccharide C atoms as a whole indicated that the monosaccharide units in **1** had the pyranose form.



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TABLE 1. Chemical Shifts of C and H Atoms of Askendoside H (**1**) and Chemical Shifts of C Atoms of Cycloorbigenin C (**2**) (δ , ppm, J/Hz, C₅D₅N, 0 = TMS for δ_C , 0 = HMDS for δ_H)

C atom	DEPT	1		2
		δ_C	δ_H	δ_C
1	CH ₂	32.46		32.74
2	CH ₂	29.67		31.40
3	CH	88.50	3.47 dd (12.3, 4.5)	78.31
4	C	42.75	–	42.42
5	CH	54.02	1.63 d (9)	53.92
6	CH	67.81	3.67 td (9, 4)	68.18
7	CH ₂	38.28		38.53
8	CH	46.77		47.09
9	C	21.31	–	21.20
10	C	29.15	–	29.53
11	CH ₂	26.24		26.26
12	CH ₂	33.04		32.95
13	C	45.78	–	45.61
14	C	46.72	–	46.79
15	CH ₂	47.86	1.53, 1.97 dd (12.9, 8)	47.68
16	CH	72.09	4.53 m	72.10
17	CH	57.38	1.70	57.40
18	CH ₃	19.66	1.30 s	18.85
19	CH ₂	30.36	0.12 d (4), 0.45 d (4)	30.23
20	CH	27.55	2.59 m	27.30
21	CH ₃	18.82	1.15 d (6.6)	20.27
22	CH ₂	38.95	2.45 dd (15, 10)	42.88
23	CH	82.77	4.55 m	73.09
24	CH	80.66	4.50 d (2.8)	79.08
25	C	72.66	–	74.29
26	CH ₃	26.35	1.52 s	24.58
27	CH ₃	28.41	1.54 s	28.91
28	CH ₃	20.08	0.88 s	20.14
29	CH ₃	28.58	1.86 s	29.32
30	CH ₃	16.22	1.28 s	16.12
<i>β-D-Xylp unit</i>				
1	CH	105.61	4.81 d (7)	
2	CH	83.62	4.01 dd (9, 6.8)	
3	CH	77.64	4.06	
4	CH	70.97	4.06 td (6.3, 2.3)	
5	CH ₂	67.01	3.50 dd (11, 9.4), 4.16 dd (11, 4.7)	
<i>α-L-Arap unit</i>				
1	CH	106.67 ^a	5.11 d (6.6)	
2	CH	73.64	4.48 dd (8.7, 6.8)	
3	CH	74.29	4.11 dd (7.5, 4.7)	
4	CH	69.14	4.20 td (3.5, 1.9)	
5	CH ₂	66.60	3.70 dd (12.2, 1.9), 4.29 dd (12.2, 3.3)	
<i>β-D-Glcp unit</i>				
1	CH	106.67 ^a	5.02 d (7.7)	
2	CH	75.89	3.87 dd (8.9, 7.7)	
3	CH	78.25	3.95 dd (9.6, 8.4)	
4	CH	72.22	4.10 dd (8.4, 7.7)	
5	CH	78.22	3.88 td (8.9, 2.4)	
6	CH ₂	63.12	4.43 dd (11.5, 2.8), 4.10 dd (12, 9)	

Resonances marked with the same letter are mutually overlapped.

The anomeric protons of D-xylose, D-glucose, and L-arabinose resonated in the PMR spectrum of **1** as doublets at δ 4.81, 5.02, and 5.11. Their SSCC ($^3J = 7, 7.7, 6.6$ Hz) also indicated that they had the pyranose form, the 4C_1 -conformation, and the β -configuration for D-xylose and D-glucose and the α -configuration for L-arabinose.

The genin, which was identified as cycloorbigenin C (**2**), was isolated from the genin part of the acid hydrolysis products [5, 6].

A comparison of ^{13}C NMR spectra of cycloorbigenin C and askendoside H showed that C-3 and C-23 of the genin and C-2 of D-xylose experienced a glycosylation effect in the latter. These resonated at δ 88.50, 82.77, and 83.62, respectively. Therefore, the new glycoside **1** was a bisdesmoside glycoside, the carbohydrate constituents of which were located on C-3 and C-23 of cycloorbigenin C and the D-glucose and L-arabinose of which were terminal.

Nuclear Overhauser Effect (NOE) difference measurements with pre-irradiation of the D-glucose anomeric proton revealed an Overhauser effect on the H-23 resonance. This determined unambiguously the location of the D-glucose on C-23. This meant that the biose located on C-3 was (α -L-arabinopyranosyl)(1 \rightarrow 2)- β -D-xylopyranose.

The difference PMR spectrum with irradiation of the D-xylose anomeric proton (δ 4.81) revealed a negative NOE on the resonance of the genin H-3 (δ 3.36) [7, 8], which confirmed the conclusion about the attachment of D-xylose to C-3.

Thus, the experimental results led to the conclusion that the new triterpene glycoside **1** was 23*R*,24*R*-cycloartan-3 β ,6 α ,16 β ,23,24,25-hexaol 3-*O*-[(α -L-arabinopyranosyl)(1 \rightarrow 2)- β -D-xylopyranoside] 23-*O*- β -D-glucopyranoside.

EXPERIMENTAL

General comments have been published [9]. The following solvent systems were used: $CHCl_3$:MeOH:H₂O (70:23:4, 1), $CHCl_3$:MeOH (10:1, 2), *n*-BuOH:Py:H₂O (6:4:3, 3). NMR spectra were recorded in Py-d₅ on a UNITYplus 400 (Varian) spectrometer. ^{13}C NMR spectra were obtained with full C–H decoupling and under DEPT conditions. 2D spectra of **1** were recorded using standard Varian programs. Chemical shifts of protons in **1** and **2** are given vs. HMDS. Chemical shifts of C atoms in ^{13}C NMR spectra of **1** and **2** are given vs. resonances of the β -C atoms of deuteropyridine (δ 123.493 vs. TMS).

Isolation and Separation of Triterpenoids from *A. taschkenticus* [10–12]. Fractions that eluted after askendoside G and contained askendoside H were combined and rechromatographed over a column using system 1 to afford **1** (170 mg, 0.0033% of air-dried raw material).

Askendoside H (1), C₄₆H₇₈O₁₉, white non-crystalline compound. Table 1 presents the PMR and ^{13}C NMR spectra of **1**.

Cycloorbigenin C (2) from 1. Askendoside H (140 mg) was hydrolyzed by methanolic H₂SO₄ (15 mL, 0.5%) on a boiling-water bath for 6 h. The mixture was diluted with H₂O. The MeOH was evaporated. The resulting precipitate was filtered off, washed with H₂O until neutral, and dried. The dry product was chromatographed over a column with elution by system 2 to afford **2** (25 mg), C₃₀H₅₂O₆, mp 256–258°C (MeOH).

PMR spectrum of **2** (400 MHz, C₅D₅N, δ , ppm, J/Hz, 0 = HMDS): 0.19 and 0.47 (d, $^2J = 4$, 2H-19), 0.89 (s, CH₃), 1.07 (d, $^3J = 6.7$, CH₃-21), 1.26, 1.27, 1.57, 1.61, 1.79 (s, 5 \times CH₃), 3.56 (dd, $^3J_1 = 11.6$, $^3J_2 = 4.7$, H-3), 3.65 (d, $^3J = 8.5$, H-24), 3.69 (td, $^3J_1 = ^3J_2 = 9.3$, $^3J_3 = 3.8$, H-6), 4.22 (td, $^3J_1 = ^3J_2 = 8.7$, $^3J_3 = 2$, H-23), 4.59 (td, $^3J_1 = ^3J_2 = 7.6$, $^3J_3 = 4.7$, H-16).

Table 1 presents the ^{13}C NMR spectrum for **2**.

The filtrate was evaporated to a small volume and heated on a water bath for 1 h to destroy methylglycosides. The solution was cooled and neutralized with BaCO₃. The precipitate was removed. The solution was evaporated to a small volume. PC using system 3 and comparison with authentic samples detected D-glucose, D-xylose, and L-arabinose. PMR and ^{13}C NMR spectra provided evidence that the aforementioned monosaccharides were present in a 1:1:1 ratio in **1**.

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